

NETRF Speaker-Cited Literature

Below find the citations of the publications made available by some of the NETRF Symposium speakers.

Session 4

Jesse Boehm (2)

Vazquez F, Boehm JS. The Cancer Dependency Map enables drug mechanism-of-action investigations. *Mol Syst Biol*. 2020;16(7):e9757. doi:10.15252/msb.20209757

Corsello SM, Nagari RT, Spangler RD, Rossen J, Kocak M, Bryan JG, Humeidi R, Peck D, Wu X, Tang AA, Wang VM, Bender SA, Lemire E, Narayan R, Montgomery P, Ben-David U, Garvie CW, Chen Y, Rees MG, Lyons NJ, McFarland JM, Wong BT, Wang L, Dumont N, O'Hearn PJ, Stefan E, Doench JG, Harrington CN, Greulich H, Meyerson M, Vazquez F, Subramanian A, Roth JA, Bittker JA, Boehm JS, Mader CC, Tsherniak A, Golub TR. Discovering the anti-cancer potential of non-oncology drugs by systematic viability profiling. *Nat Cancer*. 2020 Feb;1(2):235-248. doi: 10.1038/s43018-019-0018-6. Epub 2020 Jan 20. PMID: 32613204; PMCID: PMC7328899.

Talya Dayton (5)

Boonekamp KE, Dayton TL, Clevers H. Intestinal organoids as tools for enriching and studying specific and rare cell types: advances and future directions. *J Mol Cell Biol*. 2020 Jul 15:mjaa034. doi: 10.1093/jmcb/mjaa034. Epub ahead of print. PMID: 32667995.

Beumer, J., Puschhof, J., Bauzá-Martínez, J., Martínez-Silgado, A., Elmentaite, R., James, K. R., et al. (2020). High-Resolution mRNA and Secretome Atlas of Human Enteroendocrine Cells. *Cell*, 181(6), 1291–1306.e19.
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Gehart, H., van Es, J. H., Hamer, K., Beumer, J., Kretzschmar, K., Dekkers, J. F., et al. (2019). Identification of Enteroendocrine Regulators by Real-Time Single-Cell Differentiation Mapping. *Cell*, 176(5), 1158–1173.e16.
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Beumer, J., Artegiani, B., Post, Y., Reimann, F., Gribble, F., Nguyen, T. N., et al. (2018). Enteroendocrine cells switch hormone expression along the crypt-to-villus BMP signalling gradient. *Nature Cell Biology*, 20(8), 909–916.
<http://doi.org/10.1038/s41556-018-0143-y>

Basak, O., Beumer, J., Wiebrands, K., Seno, H., van Oudenaarden, A., & Clevers, H. (2016). Induced Quiescence of Lgr5+ Stem Cells in Intestinal Organoids Enables Differentiation of Hormone- Producing Enteroendocrine Cells. *Stem Cell*, 1–19.
<http://doi.org/10.1016/j.stem.2016.11.001>

Chris Harris (3)

Contractor, T and Harris CR (2020). Loss of copy of Hsa-mir1-2 elevates Cdk4 expression in ileal neuroendocrine tumors. *Oncogenesis* 9, 37. DOI: 10.1038/s41389-020-0221-4

Contractor, T, Clausen, R, Harris, GR, Rosenfeld, JA, Tang, L, Carpizo, DR, and Harris CR (2020). IGF2 drives formation of ileal neuroendocrine tumors in patients and in mice. *Endocr Relat. Cancer* 27, 175-186. DOI: 10.1530/ERC-19-0505.

Kobayashi, S, Contractor, T, Vosburgh, E, Tang, L, Du, N, Clausen, R, and Harris, CR (2019). Alleles of *Insm1* determine whether Rip-Tag mice produce insulinomas or nonfunctioning pancreatic neuroendocrine tumors. *Oncogenesis* 8, 16. DOI: 10.1038/s41389-019-0127-1.

Arthur Tischler (2)

Powers JF, Cochran B, Baleja JD, et al. A xenograft and cell line model of SDH-deficient pheochromocytoma derived from *Sdhb*^{+/-} rats. *Endocr Relat Cancer*. 2020;27(6):337-354. doi:10.1530/ERC-19-0474

Rai SK, Bril F, Hatch HM, et al. Targeting pheochromocytoma/paraganglioma with polyamine inhibitors. *Metabolism*. 2020;110:154297. doi:10.1016/j.metabol.2020.154297

Eugenia Xu (2)

Wong C, Tang LH, Davidson C, Vosburgh E, Chen W, Foran DJ, Notterman DA, Levine AJ, Xu EY. *Cell Death Differ*. 2020 Jan;27(1):269-283. Two well-differentiated pancreatic neuroendocrine tumor mouse models

Xu EY, Vosburgh E, Wong C, Tang LH, Notterman DA. *Oncotarget*. 2020 Jul 14;11(28):2718-2739. Genetic analysis of the cooperative tumorigenic effects of targeted deletions of tumor suppressors *Rb1*, *Trp53*, *Men1*, and *Pten* in neuroendocrine tumors in mice