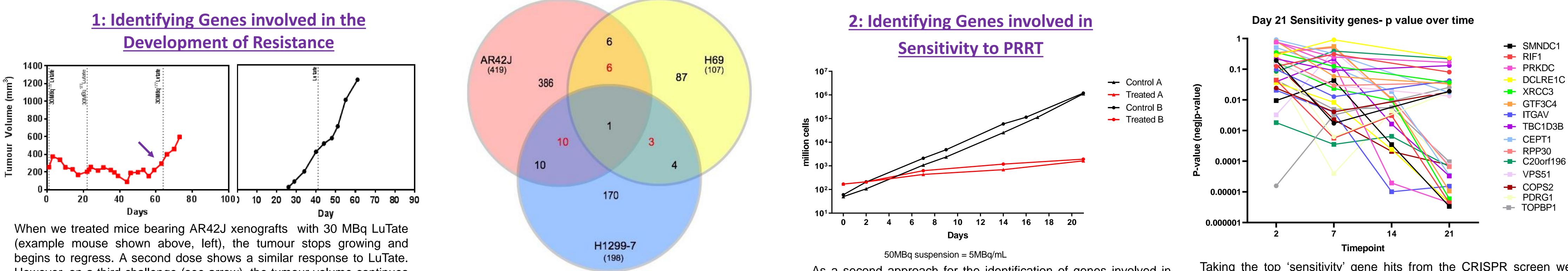
Modeling Resistance and Sensitivity to PRRT

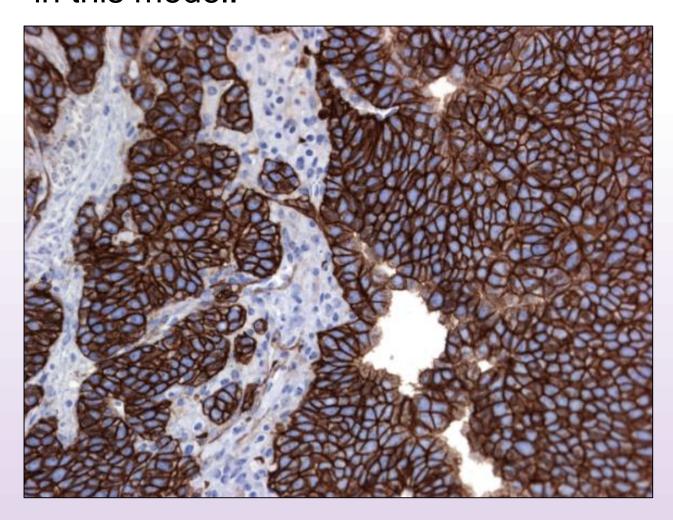
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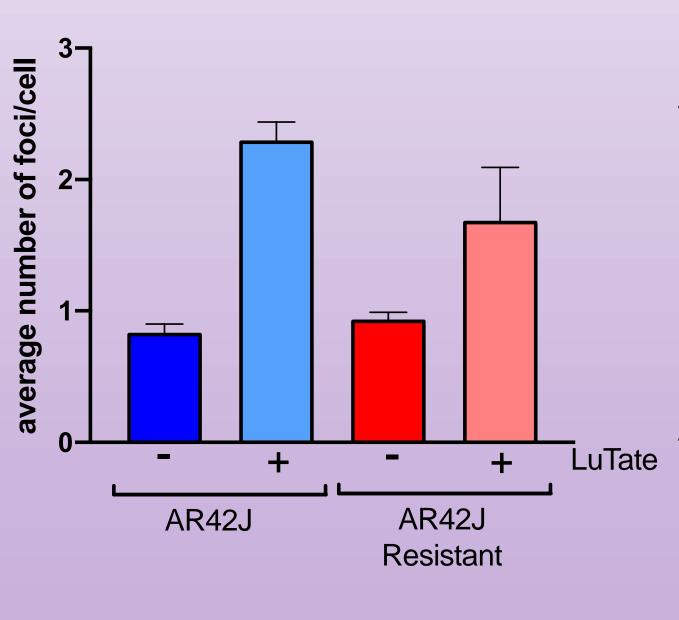
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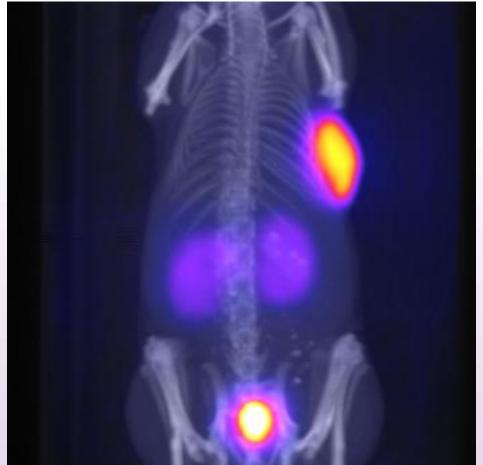
The development of resistance is a common reason for therapy failure, and ways to overcome resistance a significant area of continual research. Peptide receptor radionuclide therapy (PRRT) for neuroendocrine tumours (NET) is now routine, however cures remain rare, as some NET patients are inherently resistant to PRRT, while most develop resistance after initial success. We hypothesise that resistance to PRRT is a manifestation of a general radiation resistance phenotype, mediated through enhanced recognition and repair of radionuclide-induced DNA damage, rather than simply loss of the PRRT target (in this case, the somatostatin receptor type 2 (SSTR2). The Aims of our research are therefore:



However, on a third challenge (see arrow), the tumour volume continues to grow at the same rate. When this tumour is re-implanted into a new host and challenged with LuTate (above, right), it continues to show a resistant phenotype, and this tumour was then used to establish a 'LuTate-resistant AR42J' cell line which was then shown to also produce LuTate-resistant xenografts in subsequent hosts (data not shown). Retention of SSTR2 expression in these LuTate-resistant xenografts was confirmed by immunohistochemistry and GaTate PET imaging (below), indicating that loss of the receptor was not the mechanism of resistance in this model.







Showing that LuTate was still effective in damaging DNA in the resistant cell line, gamma-H2Ax staining in tumours 72hr after LuTate challenge showed no significant difference in the levels of accumulated DNA double-strand breaks (left). We have used this process of generating LuTate-resistant tumours and cell lines, in three cell lines, establishing a panel of tools to explore mechanisms both resistance and sensitivity, and assess drug combinations.

1: To identify genes that may contribute to the development of resistance to PRRT 2: To identify genes that play a role in sensitivity to PRRT

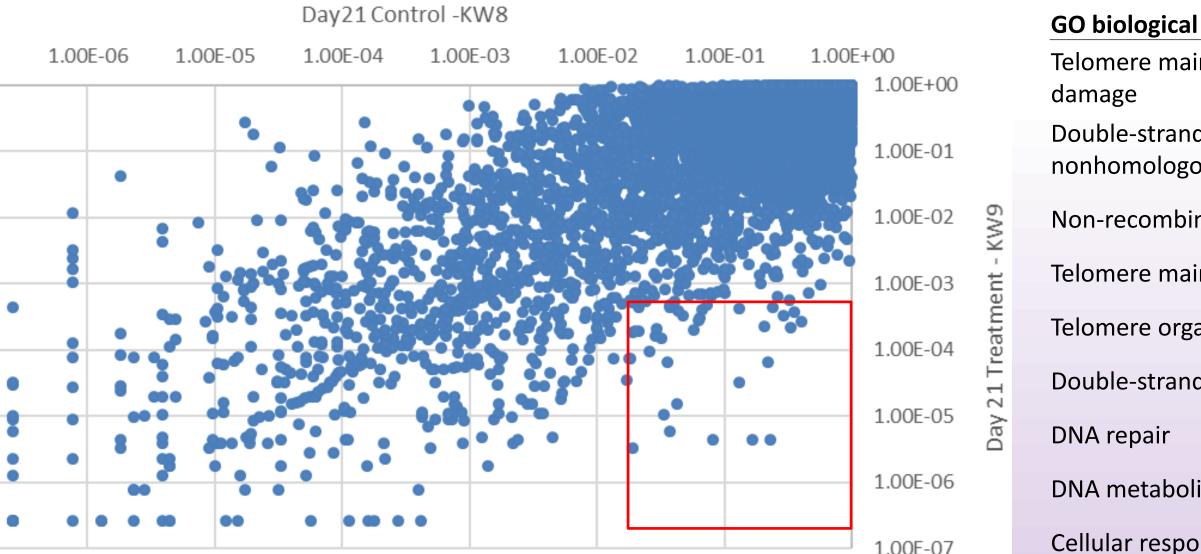
Red numbers - genes altered in same direction (either increased or decreased) Furthering the analysis of the resistant cell lines, we have examined changes to gene expression at the RNA level, through single cell RNA-sequencing technologies. Preliminary analysis of this data shows that there are 40 genes that show differential expression between their parental and resistant pairs in at least two of the models (above). When this data is expanded to look at non-significant levels of expression eight genes are identified that show a trend to similar alterations across all three resistant models. The table below shows the log-fold change in expression of these 8 genes in the resistant cell lines (as relative to their parental line), with red indicating an increase in expression and blue a decrease

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ase in expression and blue a decrease.							
	H69	H1299-7	AR42J	_			
S100A16	1.2	0.6	1.3				
GSTP1	1.1	0.5	0.4				
GADD45A	0.4	0.4	2.1				
ACTB	0.3	0.6	0.5				
HNRNPA1	-0.1	-0.5	-1.5				
SLC25A3	-0.2	-0.7	-0.6				
EEF2	-0.2	-0.5	-1.1				
EIF4B	-0.2	-0.7	-0.9				

a second approach for the identification of genes involved in th resistance and sensitivity to LuTate, we have used an biased whole genome knockout CRISPR screen. As seen ove, treatment with 5MBq/mL LuTate resulted in strong growth nibition in the H1299-7 cells. Upon sequencing, at Day 21 post eatment, we were able to identify a selection of genes that have, nen knocked out, resulted in sensitivity to LuTate (indicated by e red box in the plot below). Day 21 Treatment v control neg p-value

- Analysis is continuing in the Resistant cell lines and tumours, with **Conclusions and Future Work** the expression analysis extending to look at genes involved in the DNA damage repair pathways, and tumours being assessed Resistance to LuTate can be mediated through through Whole Genome Sequencing and further in vivo response mechanisms unrelated to SSTR2 expression. With SSTR2 experiments. expression unaltered, and gamma H2Ax foci formation in • The CRISPR screen will be repeated in a second cell line to response to LuTate unaltered in our resistant cell lines.
- further the significance of any sensitivity genes identified, with the • Eight genes have been identified as showing the same analysis extended to identifying genes involved in resistance. This trend in gene alteration in all three resistant cell lines. data will then be cross-compared with the resistant cell line data and identified targets will be validated. • The top pathway hits from the CRISPR screen, for
- sensitivity to PRRT, are all related to DNA damage repair.



• And finally, gene targets identified as resulting in sensitivity to LuTate will be used in the Resistant lines as a mechanism to hopefully overcome the LuTate resistance.

Taking the top 'sensitivity' gene hits from the CRISPR screen we have tracked the p-value of these genes over time, looking for genes that progressively result in the sensitivity observed at Day 21 (above), and then using these genes looked for any common pathways (below). As predicted in our hypothesis, pathways and genes involved in DNA damage repair were amongst the top hits for sensitivity to LuTate.

	Number of Genes from CRISPR	over/under	Fold	
l process complete	dataset	represented	Enrichment	P-value
intenance in response to DNA	2	+	> 100	3.17E-05
d break repair via				
ous end joining	3	+	69.5	1.16E-05
inational repair	3	+	63.18	1.53E-05
intenance	4	+	57.32	6.77E-07
anization	4	+	55.6	7.62E-07
d break repair	4	+	29.73	8.58E-06
	6	+	16.13	9.95E-07
lic process	6	+	11	8.91E-06
onse to DNA damage stimulus	6	+	10.58	1.11E-05

Acknowledgements

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